The Mtb Proteome Library:

Development and application of assays for targeted MS analysis of the complete proteome of *Mycobacterium tuberculosis* by SRM and SWATH-MS



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Skyline User Meeting 2013 Minneapolis





Mycobacterium tuberculosis (Mtb)



- Mycobacterium tuberculosis is the causative agent of Tuberculosis (TB)
- One third of the world's population latently infected with Mtb
- 1.7 million deaths from TB each year
- \rightarrow More efficient treatments urgently needed
- Limited availability of techniques to measure proteins with high sensitivity, selectivity and reproducibility

Traditional approach: Antibodies



Only for few targets, high cost, low throughput

Aim

To generate a resource of validated assays for the sensitive detection and accurate quantification of <u>every</u> protein of *Mycobacterium tuberculosis,* even in complex backgrounds.

The Mtb Proteome Library contains SRM assays for the entire proteome of Mtb



Schubert et al. Cell Host & Microbe 2013

The Mtb Proteome Library: A Resource of Assays to Quantify the Complete Proteome of Mycobacterium tuberculosis.

Definition of the MS-accessible Mtb proteome by discovery MS

Extensive fractionation and shotgun MS on Orbitrap XL





100% corresponds to the 4,012 annotated ORFs in Mtb (TubercuList v2.3)

RNA-seq data: Arnvig et al., PLoS Pathogens 2011

Generation of SRM assays using crude synthetic peptides



17,463 crude synthetic peptides (JPT) measured in pools of ~100 on a Qtrap 4000 in SRM-triggered MS2 mode

Increasing SRM/SWATH assay specificity and throughput by using iRTs and scheduled SRM







iRT peptides spiked into each sample allow to determine a chromatography-independent retention time (iRT) for each peptide.



Escher et al., Proteomics 2012, Using iRT, a normalized retention time for more targeted measurement of peptides

Theoretical assessment of SRM assay specificity using the SRMCollider





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| LPDGNGIELC[160]R/2,LPDG LPDGNGIELC[160]R[166]/2 LPDGNGIELC[160]R[166]/2 LPDGNGIELC[160]R[166]/2 LPDGNGIELC[160]R[166]/2 | SNGIELC[160]R,2, LPDGNGIELC[160 LPDGNGIELC[160 LPDGNGIELC[160 LPDGNGIELC[160 LPDGNGIELC[160 | 612.308612,455.2 612.308612,561.70 [R[166],2,617.312]R[166],2,617.312]R[166],2,617.312]R[166],2,617.312 | 33664, #N/A,1, y3 5658, #N/A,2, y10 2746,1041.481388, #N/A, 2746,925.454445, #N/A, 2746,592.284526, #N/A,1 2746,463.241933, #N/A,1 | 1, y9 L, y8 L, y4 L, y3 |
| LPDGNGIELC(160)R/2,LPDG LPDGNGIELC(160)R(166)/2 LPDGNGIELC(160)R(166)/2 LPDGNGIELC(160)R(166)/2 LPDGNGIELC(160)R(166)/2 LPDGNGIELC(160)R(166)/2 SSRCalc window | NGIELC [160]R,2, , LPDGNGIELC [160 , LPDGNGIELC [160 , LPDGNGIELC [160 , LPDGNGIELC [160 , LPDGNGIELC [160 | 612.308612,561.71 []R[166],2,617.312 []R[166],2,617.312 []R[166],2,617.312 []R[166],2,617.312 []R[166],2,617.312 []R[166],2,617.312 | 33664, #N/A, 2, y3 5658, #W/A, 2, y10 7746, 1041.481388, #N/A, 1 2746, 5925.454445, #N/A, 1 2746, 592.284526, #N/A, 1 2746, 463.241933, #N/A, 1 2746, 566.770714, #N/A, 2 arbitrary units | 1, y9 1, y8 1, y4 1, y3 2, y10 |
| LPDGNGTELC[160]R/2,LPDG LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 SSRCalc window | NGIELC[160]R,2, ,LPDGNGIELC[160] ,LPDGNGIELC[160] ,LPDGNGIELC[160] ,LPDGNGIELC[160] ,LPDGNGIELC[160] | 612.308612,561.71 []R[166],2,617.312]R[166],2,617.312]R[166],2,617.312]R[166],2,617.312]R[166],2,617.312]R[166],2,617.312]R[166],2,617.312 | 33664, #N/A, 2, уЗ 5658, #N/A, 2, у10 2746, 1041. 481388, #N/A, 2746, 925. 454445, #N/A, 1 2746, 592. 284526, #N/A, 1 2746, 463. 241933, #N/A, 1 2746, 566. 770714, #N/A, 2 arbitrary units | 1, y9 1, y8 1, y4 1, y3 2, y10 |
| LPDGNGTELC(160)R/2,LPDG LPDGNGTELC(160)R(166)/2 LPDGNGTELC(160)R(166)/2 LPDGNGTELC(160)R(166)/2 LPDGNGTELC(160)R(166)/2 LPDGNGTELC(160)R(166)/2 SSRCalc window Q1 mass window | NGIELC[160]R,2, , LPDENGIELC[160 , LPDENGIELC[160 , LPDENGIELC[160 , LPDENGIELC[160 , LPDENGIELC[160 | 612.308612,553.2 612.308612,551.7]R[166],2,617.311]R[166],2,617.311]R[166],2,617.311]R[166],2,617.311]R[166],2,617.311 10 0.7 | 33664, #N/A, 2, y3 565, #1, 2, y10 2746, 1041.481388, #1/A, 1746, 925.454445, #1/A, 1 2746, 592.241526, #1/A, 1 2746, 563.241933, #1/A, 1 2746, 566.770714, #1/A, 2 arbitrary units Th Th | 1,y9 1,y8 1,y4 1,y3 2,y10 |
| LPDGNGTELC(160)R/2,LPDG LPDGNGTELC(160)R(166)/2 LPDGNGTELC(160)R(166)/2 LPDGNGTELC(160)R(166)/2 LPDGNGTELC(160)R(166)/2 LPDGNGTELC(160)R(166)/2 SSRCalc window Q1 mass window Q3 mass window | NGIELC[160]R,2, , LPDONGTELC[160 , LPDONGTELC[160 , LPDONGTELC[160 , LPDONGIELC[160 , LPDONGTELC[160]] | 612.306612,563.7 [13,1166],2,617.311 [13,1166],2,617.311 [13,1166],2,617.311 [13,1166],2,617.311 [14,166],2,617.311 [10 0.7 1.0 | 33664, #N/A, 2, y3 565, #W/A, 2, y10 7746, 1041.481388, #N/A, 1 2746, 592.454445, #N/A, 1 2746, 592.284526, #N/A, 1 2746, 566.770714, #N/A, 2 arbitrary units Th Th | 1,y9 1,y8 1,y4 1,y3 2,y10 |
| LPDGNGTELC[160]#/2,LPDG LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 SSRCalc window Q1 mass window Q3 mass window Low mass threshold for transition | NGIELC [160] R, 2, , LPDONGTELC [160 , LPDONGTELC [160 , LPDONGTELC [160 , LPDONGIELC [160 , LPDONGIELC [160 , LPDONGIELC [160 | 612.30612,551.7 [12.30661,2,617.31] [12.166],2,617.31] [12.166],2,617.31] [12.166],2,617.31] [12.166],2,617.31] [10.000 | 33664, #N/A, 2, y3 565, #W/A, 2, y10 2746, 1041. 481388, #N/A, 1746, 925. 454445, #N/A, 2746, 522. 484526, #N/A, 1 2746, 566. 770714, #N/A, 2 arbitrary units Th Th Th | 1, y9 1, y8 , y4 1, y3 2, y10 |
| LPDCNGTELC[160]#/2,LPDC LPDGNGTELC[160]R[166]/2 LPDCNGTELC[160]R[166]/2 LPDCNGTELC[160]R[166]/2 LPDCNGTELC[160]R[166]/2 LPDCNGTELC[160]R[166]/2 SSRCalc window Q1 mass window Q3 mass window Low mass threshold for transition High mass threshold for transition | NGIELC [160] R.2, , LPDONGIELC [160 , LPDONGIELC [160] , LPD | 612.30612,553.7 612.30612,551.7 1]R[166],2,617.311 1]R[166],2,617.311 1]R[166],2,617.311 1]R[166],2,617.311 1]R[166],2,617.311 10 0.7 1.0 300 1500 Muchastatium basis | 33664, #N/A, 2, y3 565, #W/A, 2, y10 2746, 1041.481388, #W/A, 2746, 592.484485, #N/A, 1 2746, 592.2844526, #N/A, 1 2746, 566.770714, #N/A, 2 arbitrary units Th Th Th | 1, y9 , y8 , y4 , y4 , y3 2, y10 |
| LPDENGTELC [160] R/2, LPDC LPDGNGTELC [160] R(166)/2 LPDGNGTELC [160] R(166)/2 LPDENGTELC [160] R(166)/2 LPDENGTELC [160] R(166)/2 LPDENGTELC [160] R(166)/2 SSRCalc window Q1 mass window Q3 mass window Low mass threshold for transition High mass threshold for transition Genome Consider isotopas up to | NGIELC [160] R.2, , LPDENGIELC [160 , LPDENGIELC [160 , LPDENGIELC [160 , LPDENGIELC [160 , LPDENGIELC [160]]]]]]]]]]]]]]]]]]] | 612.30612,553.7 612.306612,551.7 1]R [166],2,617.311 1]R [166],2,617.311 1]R [166],2,617.311 1]R [166],2,617.311 1]R [166],2,617.311 10 0.7 1.0 300 1500 Mycobacterium bovis | 33664, #N/A, 2, y3 555, #W/A, 2, y10 7746, 1041.481388, #W/A, 1 2746, 592.284526, #N/A, 1 2746, 592.284526, #N/A, 1 2746, 566.770714, #N/A, 2 arbitrary units Th Th Th Th | 1, y9 , y8 , y4 , y4 , y3 2, y10 |
| LPDGNGTELC[160]#/2,LPDG LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 LPDGNGTELC[160]R[166]/2 SSRCalc window Q1 mass window Q3 mass window Low mass threshold for transition High mass threshold for transition Genome Consider isotopes up to | INGIELC [160] R, 2, , , LPDENGTELC [160 , LPDENGTELC [160] , | 612.30612,553.7 612.306612,561.7]R[166],2,617.311]R[166],2,617.311]R[166],2,617.312]R[166],2,617.312]R[166],2,617.312]R[166],2,617.312 10 0.7 1.0 300 1500 Mycobacterium bovis 3 1 | 33664, #N/A, 2, y3 5658, #W/A, 2, y10 7746, 1041.481388, #W/A, 1 7746, 592.284526, #N/A, 1 7746, 562.284526, #N/A, 1 7746, 566.770714, #N/A, 2 arbitrary units Th Th Th Th Th amu | 1, y9 1, y8 1, y4 1, y3 2, y10 |

Validation of the Mtb Proteome Library in unfractionated whole cell lysates by SRM





To validate all these SRM assays, over 200 scheduled SRM runs were needed.

Reiter et al., Nature Methods 2011, mProphet: automated data processing and statistical validation for large-scale SRM experiments

The Mtb Proteome Library contains SRM assays for the entire proteome of Mtb



Schubert et al. Cell Host & Microbe 2013

The Mtb Proteome Library: A Resource of Assays to Quantify the Complete Proteome of Mycobacterium tuberculosis.

The Mtb Proteome Library is a publicly available resource of MS reference data, SRM assays and their validation

www.PeptideAtlas.org/PASSEL

Search All Builds Current Build Queries SRMAtlas PTPAtlas Submission Peptide Protein Search All Builds Current Build Queries SRMAtlas PTPAtlas Submission SRMATLAS HOME PeptideAtlas Build: M. tuberculosis 2013-05 Query Transitions Transition Lists SRMAtlas Builds PASSEL Experiments PASSEL Data DATA ACCESS Protein Name: Rv0001 (00) Search SRM Assa View SRM Builds Works best under Firefox Get SRM Experiment Transitions Gene Name dnaA BACKGROUND Project Home Rv0001ldna Identification Status canonical Form Resultset ChromaVis Plot Data Contributor Publications ProteinProphet Probability 1.00000 External Links Mult Hyp Testing Probability lect left and right chromatograms from the Results tab Contacts Distinct Peptides SRM/MRM Assays Drag the red area over the context panel to shift the focus panel. When the cursor is a crosshair, you may draw a new context window. Lock/unlock X and Y axes as desired Total Observations ProteinProphet-adjusted N Obr SRM/MRM Glossar 126 ormalized PSMs per 100K YVSTEEFTNDFINSLR (1933.906 Daltons) +2 . light YVSTEEFTNDFINSLR (1933,906 Daltons) +3 . light LOGIN Experiment: M. tuberculosis SRM atlas (Schubert et al Spectrum file: olgas_H110902_105_TRID30.mzXML Experiment: M. tuberculosis SRM atlas (Schubert et al Spectrum file: olgas_H110902_107_TRID31.mzXML 0 Sequence Position Chromatogram ID: 296421 Chromatogram ID: 296463 mQuest: best pg RT=27.726 S/N=66.713 log max apex intens=4.564 nQuest: best pg RT=27.617 S/N=23.614 log max apex intens=3.98 y3 y4 y5 y6 y8 10.000 y4 y6 y7 y8^2 OTHER RESOURCES 8,000 30,000 2222 6.000 Sequence Coverage 20,000 Unlikely (theoretical) 0 00 Systems 🚯 4,000 Biology 10.000 2.00 erved peptide with single genome map rotein coverage by observed peptides tides unlikely to be observed Sequence Display Mode: Tryptic SGFTTVWNAVVSELNGDPK VI QPLTIVEOFALAR GONQHENPSYFIT TTTDNDEIDDEAAAR GONQHENPSYFIT EAAGNTAQR LFPGRR VK IVSTEEFINT CONTROL OLATLEDR LR TR LGK TELLE Y 🔒 EVK ELTTR IN OF R R RVF www.SRMAtlas.org Protein Coverage = 62.1% (75.9% of likely observable sequence) Auto Scale Y Axis nt Build Queries SRMAtlas PTPAtla Transition Lists SRMAtlas Builds PASSEL Experiments PASSEL Data Query Transitions form results TSV thg ion Rank RI SSRT External Links B-0001 YTEDTEVIGASNE QTOF 745.87 2 504,252 1 7771 33.9 1 73 1 v5 By0001 R YTEDTEVIGASNE 1.73 OTOF 745.87 2 617,3365 1 v6 2 6015 33.9 1 By0001 B YTEDTEVIGASNB F 1.73 OTOF 745.87 2 716,4049 1 y7 3 5012 33.9 1 R YTEDTEVIGASNE QTOF 745.87 Bv0001 1.73 2 1226.6164 1 v11 4 4579 33.9 LOGIN Rv0001 K VDDGPSSDANLSAPLTPQQR A 1.68 QTOF 1034.50 2 839.4734 1 y7 1 7796 24.9 Rv0001 K VDDGPSSDANLSAPLTPOOR A 1.68 QTOF 1034.50 2 528,2889 1 v4 2 5680 24.9 1 Rv0001 K VDDGPSSDANLSAPLTPQQR A 1.68 QTOF 1034.50 2 997.5425 1 y9 3 2548 24.9 1 Rv0001 K VDDGPSSDANLSAPLTPQQR A 1.68 QTOF 1034.50 2 910,5105 1 v8 4 2217 24.9 1 Rv0001 K **YVSTEEFTNDFINSLR** D 1.37 QTOF 967.96 2 749.4304 1 y6 1 2722 37.4 1

www.PeptideAtlas.org

YVSTEEFTNDFINSLR

Rv0001 K YVSTEEFTNDFINSLR

By0001 K YVSTEEETNDEINSLB

BV0001 B FAHAAAIAIAFAPAB

Rv0001 R FAHAAALAIAEAPAR

R FAHAAALAIAFAPAR

THLLHAAGNYAQR

THLLHAAGNYAGR

THLLHAAGNYAQR

K THULHAAGNYAOR

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Systems 🔒

Biology

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D 1.37

D 1.37

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A 1.70 OTOF-pred 493.94

A 1.70 QTOF-pred 493.94

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1.57 QTOF-pred 363.69

A 1.70 QTOF-pred 493.94

QTOF 967.96

QTOF 967.96

OTOF 967.96

2 1079.5480

2 489,2780

2 263 1390

3 219 11

3 614.33

3 1234.66

3 569.28

4 374.22

4 239.11

4 352.20

4 915.48

1 y9

1 v4

1 h2

1 h2

1 v6

1 b13

1 b6

1 y3

1 b2

1 b3

1 b9

2 1623 37.4 1

3 1359 37.4 1

4 1298 37.4 1

1 100 29 3 1

2 81 29.3 1

3 74 29.3

4 58 29.3 1

1 70 21.4 1

2 43 21.4 1

3 36 21.4 1

4 36 21.4 1

Application of the Mtb Proteome Library to study the dynamics of the DosR regulon of Mtb under hypoxia

Mycobacterium bovis BCG Re-aeration No aeration Culture density (OD₆₀₀) -5 -4 -3 -2 🖌 2 3 4 5 6 7 8 9 10 11 Time (days) Re-aeration change Fold 0.5 0.25 6h Time (days)

| DosR study summary | | |
|---|----|----------|
| Total number of proteins | 53 | † |
| Proteins with SRM assays | 52 | |
| Proteins with validated assays in exponential and stationary growth phase | 37 | Skyline |
| Proteins detected in DosR study | 45 | |



Application of the Mtb Proteome Library to study the dynamics of the DosR regulon of Mtb under hypoxia



| DosR study summary |
|---|
| Total number of proteins |
| Proteins with SRM assays |
| Proteins with validated assays in exponential and stationary growth phase |



Proteins detected in DosR study



Statistical analysis of SRM data by SRMstats



SRMstats

Protein significance analysis in SRM measurements

Goal

A statistical framework is proposed for protein quantification in SRM experiments based on a family of linear mixed-effects models. The framework is sensitive and flexible, and is applicable to a variety of experimental designs and to both label-based and label-free workflows.







Chang et al., Mol Cell Proteomics 2012. Protein significance analysis in SRM measurements

Absolute label-free quantification by SRM exploits the linear correlation of the sum of the top transitions of the top peptides per protein and the protein concentration



Linear correlation established using 34 anchor proteins quantified by AQUA peptides MS intensity: sum of 2 most intense transitions of the 3 most intense peptides per protein

Ludwig et al., MCP 2011, Estimation of absolute protein quantities of unlabeled samples by SRM

Proteome-wide absolute abundance estimates for Mtb



Proteins

SWATH-MS: Data-independent acquisition with targeted data extraction



Gillet et al., MCP 2012, Targeted data extraction of the MS/MS spectra generated by data-independent acquisition: a new concept for consistent and accurate proteome analysis.

Figure by Christina Ludwig

Generation of the Mtb SWATH library





SWATH-MS allows reproducible, high proteome coverage measurements of Mtb in a single run





Summary



- The Mtb Proteome Library is a public resource of SRM assays for the entire proteome of Mtb
 - SRM assays generated from crude synthetic peptides and validated in whole cell lysates
 - Data analysis with **Skyline** and supporting tools: iRT peptides, SRMCollider, mProphet, SRMstats
 - Data can be browsed on and downloaded from www.SRMAtlas.org, www.PeptideAtlas.org/passel
- Application of the Mtb Proteome Library to study the dynamics of the DosR regulon of Mtb under hypoxia
- Absolute label-free quantification by SRM exploits the linear correlation of the sum of the top transitions of the top peptides per protein and its absolute concentration.
- Expansion of the Mtb Proteome Library for use with SWATH-MS
- SWATH-MS allows reproducible, high proteome coverage measurements of Mtb in a single run

ETH Zurich

- o Prof. Ruedi Aebersold
- Christina Ludwig
- o Jeppe Mouritsen
- George Rosenberger
- Hannes Röst (Mon 5:45 pm and WP 595, Wed)
- Ludovic Gillet (TOD pm, Tue 5:30 pm)
- o Ben Collins (WP 688, Wed)
- o Alessio Maiolica, Mariette Matondo

University of Ghana

o Dr. Patrick K. Arthur

Max Planck Institute Berlin

- o Prof. Stefan Kaufmann
- o Dr. Martin Gengenbacher

ISB Seattle

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- o Terry Farrah
- Samuel Bader (ABSciex, Mon 7 am)

University of Washington

o Brendan MacLean (TP 499, Tue)

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